



Review

Melanocortin MC₁ receptor in human genetics and model systems

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ABSTRACT

The melanocortin MC₁ receptor is a G-protein coupled receptor expressed in the melanocytes of the skin and hair and is known for its key role in the regulation of human pigmentation. Melanocortin MC₁ receptor activation after ultraviolet radiation exposure results in a switch from the red/yellow pheomelanin to the brown/black eumelanin pigment synthesis within cutaneous melanocytes; this pigment is then transferred to the surrounding keratinocytes of the skin. The increase in melanin maturation and uptake results in tanning of the skin, providing a physical protection of skin cells from ultraviolet radiation induced DNA damage. Melanocortin MC₁ receptor polymorphism is widespread within the Caucasian population and some variant alleles are associated with red hair colour, fair skin, poor tanning and increased risk of skin cancer. Here we will discuss the use of mouse coat colour models, human genetic association studies, and *in vitro* cell culture studies to determine the complex functions of the melanocortin MC₁ receptor and the molecular mechanisms underlying the association between melanocortin MC₁ receptor variant alleles and the red hair colour phenotype. Recent research indicates that melanocortin MC₁ receptor has many non-pigmentary functions, and that the increased risk of skin cancer conferred by melanocortin MC₁ receptor variant alleles is to some extent independent of pigmentation phenotypes. The use of new transgenic mouse models, the study of novel melanocortin MC₁ receptor response genes and the use of more advanced human skin models such as 3D skin reconstruction may provide key elements in understanding the pharmacogenetics of human melanocortin MC₁ receptor polymorphism.

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1. Introduction

The melanocortin MC₁ receptor is a seven trans-membrane G-protein coupled receptor expressed in the melanocytes of the skin and hair follicles, but absent from melanocytes in the uveal tract of the

eye. Activation of melanocortin MC₁ receptor by ultraviolet radiation increases synthesis of the dark eumelanin pigment, resulting in the visible darkening of the skin known as the tanning response. Melanocortin MC₁ receptor stimulation also results in increased melanocyte dendricity, proliferation, cell survival and DNA repair [for a detailed review of melanocortin MC₁ receptor function see Abdel-Malek et al. (2010, 2008); Beaumont et al. (2009); Garcia-Borron et al. (2005)]. Loss of melanocortin MC₁ receptor function results in synthesis of the red/yellow pheomelanin pigment by

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melanocytes, giving rise to red hair colour in humans or yellow coat colour in mice.

Melanocortin MC₁ receptor was one of the first genes to be associated with normal human pigmentation variation, and is the most important predictor of skin and hair colour within the Caucasian population. Genetic association studies have identified melanocortin MC₁ receptor polymorphisms that are associated with red hair, fair skin, poor tanning, freckling and increased skin cancer risk; this is termed the red hair colour phenotype. Here we describe the use of genetic association studies to determine the penetrance and interaction of several common variant melanocortin MC₁ receptor alleles found in the Brisbane twin nevus study, the use of melanoma and non-melanocytic transfected cell lines to analyse melanocortin MC₁ receptor wild type and variant allele function, as well as the use of genotyped melanocyte strains in monoculture and co-culture, a more physiologically intact system. We will also discuss other more complex systems for further elucidation of the role of melanocortin MC₁ receptor in the regulation of pigmentation and the response to ultraviolet radiation, such as the use of 3D skin reconstruction and transgenic mouse models.

Intense research into melanocortin MC₁ receptor function using human and mouse genetics and *in vitro* model systems has revealed a network of interactions involved in the upstream regulation and downstream induced signalling pathways. However, recent research has posed new questions about key components involved in the melanocortin MC₁ receptor response. Fig. 1 summarises the different ligands, signalling pathways and responses involved in melanocortin MC₁ receptor activation. The canonical pathway after melanocortin MC₁ receptor binding by the α -melanocyte stimulating hormone agonist, produced in response to ultraviolet radiation-induced DNA damage within the surrounding keratinocytes, involves a cAMP signalling cascade. This ultimately results in upregulation of the microphthalmia transcription factor, a master regulator of melanocyte cell differentiation. Microphthalmia transcription factor controls the

transcription of many important pigment genes such as tyrosinase, the main enzyme responsible for melanin synthesis.

Recently, a novel melanocortin MC₁ receptor ligand has been discovered. Black coat colour of dogs was found to be due to an activating mutation in an antibacterial β -defensin gene, CBD103 (Candille et al., 2007). This gene also induced a black coat colour in transgenic mice, and can bind to mouse melanocortin MC₁ receptor, but does not activate cAMP. The human equivalent, human β -defensin 3, was also found to bind to melanocortin MC₁ receptor. Human β -defensin 3 is also produced by keratinocytes in response to ultraviolet radiation (Glaser et al., 2009). Further research is needed to determine if human β -defensin 3 acts as a melanocortin MC₁ receptor agonist by binding and stimulating an alternative, non-cAMP dependent pathway.

After ultraviolet radiation exposure of the skin, keratinocytes can produce reactive oxygen species that are detected and transferred to melanocytes by passive diffusion (Pelle et al., 2005) and absorbed by the melanin pigments they produce. Moreover, α -melanocyte stimulating hormone treatment has been found to protect melanocytes from reactive oxygen species by induction of catalase enzyme activity (Song et al., 2009) or stimulating intracellular trafficking of the catalase protein to the cell periphery (Maresca et al., 2010).

Other non-pigmentary actions of melanocortin MC₁ receptor may be mediated through early response genes such as the nuclear hormone receptor NR4A family, which are involved in DNA repair (Smith et al., 2008). Dopachrome tautomerase, previously thought to be primarily involved in melanin synthesis, may also play a role in protection from DNA damage (Michard et al., 2008) and cell survival (Sendoel et al., 2010).

2. Melanocortin MC₁ receptor genetic association studies with pigmentation phenotypes

In the late 20th century the role of melanocortin MC₁ receptor in melanogenesis was discovered largely due to investigations into the genetics of mouse coat colour, as well as studies on cultured melanocytes and melanoma cells, and the notable effects of the melanocortin MC₁ receptor agonist α -melanocyte stimulating hormone on skin colour in humans. The association between melanocortin MC₁ receptor variant alleles and red hair in humans was first described by Valverde et al. (1995), where they employed direct sequencing of the melanocortin MC₁ receptor gene in a group of unrelated Caucasian volunteers, some of whom were chosen for extremes of hair colour. More detailed studies utilising a large adolescent twin collection in Australia (Brisbane twin nevus study), revealed the relatively common high penetrance melanocortin MC₁ receptor variant *R* alleles responsible for the red hair colour phenotype in Caucasians. These variant alleles include D84E, R151C, R160W and D294H (Box et al., 1997; Duffy et al., 2004). Low penetrance variant *r* alleles included V60L, V92M and R163Q. Two other melanocortin MC₁ receptor alleles, R142H and I155T, did not show a statistically significant association with red hair, probably due to their relatively low frequencies in the population (Duffy et al., 2004), although I155T was significantly associated with blonde hair.

R142H and I155T have both been found to show association with red hair colour in familial studies (Box et al., 1997; Flanagan et al., 2000). The association of R142H with red hair colour was confirmed by a recent meta-analysis, and given the odds ratios were similar to the other high penetrance *R* variants (Raimondi et al., 2008), and *in vitro* studies have found a functional impairment similar to the other *R* variant receptors (Beaumont et al., 2007), we thus classify R142H as a high penetrance *R* variant allele. Although I155T was initially classified as a familial *R* allele (Beaumont et al., 2005), given the lack of significant association with red hair colour in the meta-analysis (Raimondi et al., 2008), and the relatively small functional impairment of the I155T receptor compared to the other *R* variant receptors

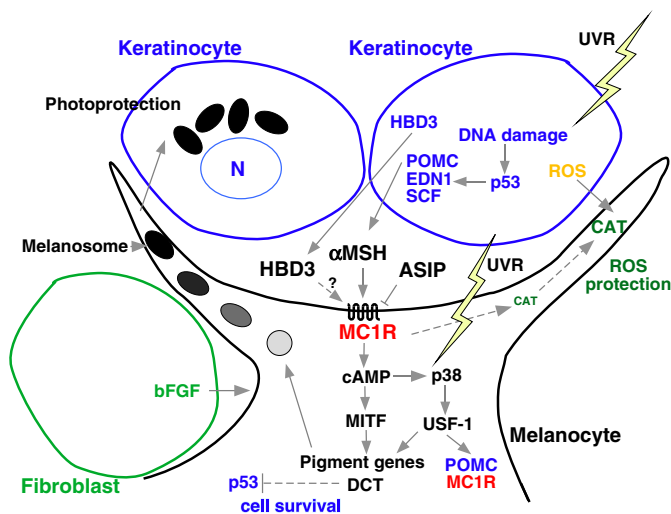


Fig. 1. The melanocyte and the tanning response. Adapted from Beaumont et al. (2009). Exposure of human skin to ultraviolet radiation results in the release of various factors by keratinocytes and melanocytes. Proopiomelanocortin (POMC) is converted to α -melanocyte stimulating hormone (α -MSH), which stimulates melanocortin MC₁ receptor (MC₁R) cAMP signalling in the melanocyte causing the upregulation of genes involved in melanogenesis, ultimately resulting in the tanning response. Agouti signaling protein (ASIP) antagonises the action of α -MSH. Dotted lines and ? indicate interactions that are in need of further research. ROS = reactive oxygen species, UVR = ultraviolet radiation, HBD3 = human β -defensin 3, EDN1 = endothelin 1, SCF = stem cell factor, CAT = catalase, bFGF = basic fibroblast growth factor, MITF = microphthalmia transcription factor, DCT = dopachrome tautomerase, USF-1 = upstream transcription factor 1.

Table 1
MC1R variant alleles in hair and skin colour.

MC1R allele		Red hair %	Blonde hair %	L brown hair %	D brown hair %	Black hair %	Fair skin %
Wild type	+/+	0	10.4	34.7	48.6	6.3	21
+/V60L	+/r	0.4	14.6	34.8	44.7	5.5	24
V60L/V60L	r/r	2.5	17.5	32.5	47.5	0	50
+/D84E	+/R	0	21.7	34.8	43.5	0	64
D84E/D84E	R/R	–	–	–	–	–	–
+/V92M	+/r	1.4	15.9	35.9	42.3	4.6	39
V92M/V92M	r/r	0	0	20	70	10	43
+/R142H	+/R	0	15.4	15.4	61.5	7.7	10
R142H/R142H	R/R	–	–	–	–	–	–
+/R151C	+/R	0.9	23.5	42.6	30.9	2.2	45
R151C/R151C	R/R	74.1	14.8	3.7	7.4	0	94
+/I155T	+/r	0	24	36	40	0	31
I155T/I155T*	r/r	0	0	0	100	0	0
+/R160W	+/R	4	14.6	45.5	34.8	1.1	44
R160W/R160W	R/R	57.1	14.3	14.3	14.3	0	50
+/R163Q	+/r	0	16	40	42	2	24
R163Q/R163Q	r/r	0	0	50	25	25	50
+/D294H	+/R	0	14	34	48	4	40
D294H/D294H*	R/R	100	0	0	0	0	100

Adapted from data in Beaumont et al. (2007). *For these genotypes data was obtained from only one individual. R = high penetrance allele, r = low penetrance allele, r = familial association with red hair colour phenotype/functionally classified as equivalent to r. L brown = Light brown, D brown = Dark brown.

[Beaumont et al. (2009) and see discussion below], here we classify I155T as a potential *r* allele (Table 1). Although interestingly, the meta-analysis did find that the I155T allele was significantly associated with melanoma (Raimondi et al., 2008). An increasing number of studies have found that the association of melanocortin MC₁ receptor variant alleles with skin cancer risk is independent of pigmentation phenotype, demonstrating melanocortin MC₁ receptor has non-pigmentary functions (Bastiaens et al., 2001; Box et al., 2001; Kanetsky et al., 2010; Landi et al., 2005; Liboutet et al., 2006). However, given that gaps exist in our understanding of the melanogenic intermediates and the process of melanin polymerization, there remains the possibility of unknown melanocortin MC₁ receptor mediated pigmentary responses being involved.

In order to look more specifically at individual variant alleles, we have separated out effects of melanocortin MC₁ receptor variants in the +/R or +/r heterozygous and R/R, or r/r homozygous states as shown in Table 1 [adapted from Beaumont et al. (2007)], this allows consideration of melanocortin MC₁ receptor alleles without potential transcomplementation effects. We have also removed the one red haired individual who was initially genotyped as wild type, as we later sequenced the melanocortin MC₁ receptor gene to find this individual was compound heterozygous for two rare frameshift mutations (Beaumont et al., 2008).

Although red hair colour is generally inherited in a recessive manner, the +/R heterozygote effect of the common melanocortin MC₁ receptor alleles can be seen by increases in the percentage fair skinned individuals, as well as increases in the percentage of red and particularly blonde hair. This correlates with data obtained from our *in vitro* studies on dominant negative effects of melanocortin MC₁ receptor variant receptors on the wild type melanocortin MC₁ receptor (Beaumont et al., 2007). The only variant that did not show a lightening effect on skin and hair colour in the heterozygous state was the R142H allele. This was the only variant that did not show a dominant negative effect on the wild type receptor *in vitro* (Beaumont et al., 2007).

Unfortunately, due to their rarity we were unable to obtain a large number of some R/R homozygous genotypes such as R142H, I155T and D294H, however the strong effect of the R160W and particularly the R151C variants on hair and skin colour in the homozygous state was confirmed, while the V60L had the largest effect of the *r/r* variants, which again agrees with our *in vitro* studies.

3. Melanocortin MC₁ receptor *in vitro* studies in melanoma and non-melanocytic cell lines

Given their association with a loss of eumelanin pigment, it was assumed that melanocortin MC₁ receptor variant alleles would be loss of function receptors. However, it was clear from the genetic studies that there were some differences in the strength of penetrance between different alleles. Due to difficulties culturing primary melanocytes and obtaining the different melanocortin MC₁ receptor variant strains, many *in vitro* studies on melanocortin MC₁ receptor function have been performed in melanoma or non-melanocytic cell lines, transfected with melanocortin MC₁ receptor wild type or variant alleles. The use of expression vector transfected cell lines also removes the influence of different genetic backgrounds and levels of melanocortin MC₁ receptor protein expression found between melanocyte strains.

The assay of cAMP after stimulation with melanocortin MC₁ receptor agonists such as the super-potent α -melanocyte stimulating hormone analogue [Nle⁴, D-Phe⁷] α -melanocyte stimulating hormone (NDP-MSH) has routinely been used as a measure of melanocortin MC₁ receptor function. We have systematically studied the cellular localisation and function of melanocortin MC₁ receptor variant alleles in transiently transfected melanoma cells, as well as stably transfected HEK293 cells. Notably, we discovered that a number of melanocortin MC₁ receptor variant receptors were intracellularly retained, thus resulting in a corresponding reduction in cAMP signalling (Beaumont et al., 2009, 2005, 2007).

We have also determined that the *in vitro* functional ability of variant receptors is similar to the strength of the reported genetic associations. The relative cAMP signalling abilities of the common melanocortin MC₁ receptor variant alleles in our assays were as follows: V92M \geq wild type > R163Q > I155T > V60L \approx R160W > R151C \approx D84E \approx R142H > D294H [Beaumont et al. (2009, 2007) and see Table 1].

All variant receptors showed some decrease in cAMP signalling compared to the wild type receptor, with the exception of the V92M variant low penetrance *r* allele. It remains to be seen whether the V92M variant displays altered function in some other way such as internalisation, desensitisation or non-cAMP mediated signalling. Some reports have noted a reduced affinity of the V92M variant for α -melanocyte stimulating hormone (Xu et al., 1996), however we did not see altered affinity in our studies.

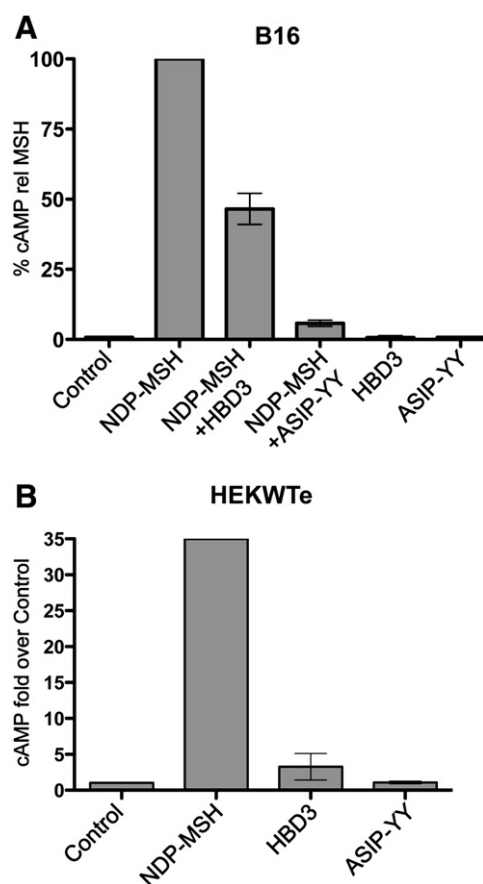


Fig. 2. cAMP accumulation in B16 or HEK293 cells stably transfected with melanocortin MC₁ receptor (HEKWTc). Cells were incubated in serum-free media for at least 2 h. Cells were pre-incubated with 0.1 mM IBMX for 15 min, then stimulated with the indicated ligands for 10 min. cAMP levels were quantified with the cAMP EIA system (Amersham Biosciences). The bars indicate the range from two independent experiments. A) 0.5 nM or 1 nM NDP-MSH alone or in combination with 100 nM agouti signaling protein peptide (ASIP-YY) (McNulty et al., 2005) or HBD3, or 100 nM ASIP-YY or HBD3 alone. Data was normalised to NDP-MSH, which was set at 100%. Results are from two independent experiments. B) 1 nM NDP-MSH, 100 nM HBD3 or 100 nM ASIP-YY. Data was expressed as fold over the control. Results are from 2 independent experiments, except for NDP-MSH, which is from only one experiment.

Although the α -melanocyte stimulating hormone induced signalling pathways have been well studied, the functional effects of the novel melanocortin MC₁ receptor ligand human β -defensin 3 [also known as HBD3, defensin, beta 103A (DEFB103A)] are as yet unknown. For the purpose of characterising the signalling pathways involved, we have used stably transfected HEK293 cell lines and B16 cells. B16 mouse melanoma is another commonly used cell line in the study of melanocortin MC₁ receptor. Unlike many human melanoma cell lines, B16 endogenously express mouse melanocortin MC₁ receptor and retain the ability to respond to α -melanocyte stimulating hormone.

Our preliminary data confirms that human β -defensin 3 does not activate cAMP signalling in B16 (Fig. 2A) or melanocortin MC₁ receptor expressing HEK293 cells (Fig. 2B), and that human β -defensin 3 can compete with NDP-MSH to inhibit melanocortin MC₁ receptor-induced cAMP signalling, although not to the same extent as the known melanocortin MC₁ receptor antagonist agouti signaling protein (Fig. 2A). This is probably due to the lower affinity of the human β -defensin 3 ligand for melanocortin MC₁ receptor compared to NDP-MSH or agouti signaling protein. Human β -defensin 3 binding competition studies with α -melanocyte stimulating hormone have been published previously (Candille et al., 2007), although binding of human β -defensin 3 to mouse melanocortin MC₁ receptor was not

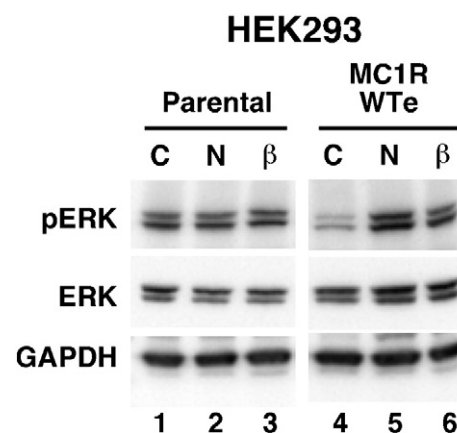


Fig. 3. ERK phosphorylation in response to α -melanocyte stimulating hormone and human β -Defensin 3. Lanes are as indicated C = Control, N = 10 nM NDP-MSH, β = 10 nM HBD3. HEK293 untransfected cells (parental) or HEK293 stably expressing melanocortin MC₁ receptor (WTe) were pre-incubated in serum-free media for at least 2 h, then stimulated with the indicated ligands for 5 min. Western immuno-blotting with the indicated antibodies was performed using total cell extracts. pERK (Cell Signalling) is specific to the phosphorylated Thr202/Tyr204 sites of ERK1/2. ERK (Cell Signalling) recognises all forms of the ERK protein. Anti-GAPDH (R&D Systems) was used as a loading control. This blot is representative of three independent experiments.

determined. Note that HEK293 cells transfected with the empty vector pcDNA3.1 did not respond to any of the melanocortin MC₁ receptor ligands (data not shown), confirming the specificity of the ligands for melanocortin MC₁ receptor.

We have also looked at melanocortin MC₁ receptor induced mitogen-activated protein kinase (MAPK) activation, as an alternative to the traditional cAMP pathway. Interestingly, although melanocortin MC₁ receptor-induced MAPK signalling is thought to be activated via crosstalk with cAMP (Busca et al., 2000), our preliminary data indicates that despite being unable to activate cAMP (Fig. 2) human β -defensin 3 may activate ERK1/2 in HEK293 cells transfected with melanocortin MC₁ receptor (Fig. 3, lane 6). MAPK may be responsible for the proliferative effect of melanocortin MC₁ receptor activation (Dumaz and Marais, 2005), ERK also phosphorylates microphthalmia transcription factor (Hemesath et al., 1998). The way in which the human β -defensin 3 melanocortin MC₁ receptor ligand is able to induce melanogenesis – at least in dogs and mouse models – is still in need of further research.

4. Genotyped primary melanocytes in monoculture and co-culture

Although initially there was some difficulty demonstrating melanocortin MC₁ receptor-induced melanogenesis using *in vitro* cultured melanocytes, by changing culture conditions, the use of human melanocytes of defined melanocortin MC₁ receptor genotype has provided a major experimental approach for our laboratory and others to define the functional consequences for each melanocortin MC₁ receptor allele as, under the appropriate growth conditions and short-term passage, melanocytes can continue to express the pigimentary phenotype characteristic of the skin type from which they were derived (Leonard et al., 2003). Clonal melanocyte cultures allow large amounts of a single cell type to be prepared for biochemical, cellular, molecular and genetic analysis and have facilitated investigation of melanocyte differentiation, and responses to growth factors, hormones and ultraviolet radiation (Kadekaro et al., 2006). However, despite maintaining their *ex-vivo* skin pigmentation phenotype, melanocytes in culture do display different morphological and phenotypic characteristics to that of cells seen *in situ*. They grow *in vitro* with a bipolar morphology and acquire expression of several melanoma-associated antigens (Hsu et al., 2002). This suggests an important role of the microenvironment in controlling the phenotype

of melanocytes. In normal skin, melanocytes are intimately associated with neighbouring keratinocytes forming the epidermal-melanin unit, which allows not only the transfer of melanin-containing melanosomes into keratinocytes, but also extensive interactions. Upon co-culture with keratinocytes, melanocytes regain a multi-dendritic differentiated phenotype resembling that seen *in vivo* (Roberts et al., 2008). Moreover, the defined serum-free keratinocyte medium used in co-culture permits the study of melanocortin MC₁ receptor signalling in medium lacking exogenous cAMP elevating agents normally required for melanocyte monoculture (Halaban, 2000).

Melanocytes in this environment also have a repertoire of signalling pathways activated by keratinocyte-derived factors that may better resemble the situation *in vivo* (Cook et al., 2003). This is an important aspect for studies on melanocortin MC₁ receptor function in light of the considerable cross-talk between different melanocyte signal transduction cascades (Fig. 1). Such pathway interactions may underlie the important role of keratinocytes in the regulation of dendricity, the proportions of pheomelanin and eumelanin, degree of ultraviolet-B stimulated eumelanogenesis, and require consideration.

We have employed the melanocyte:keratinocyte co-culture system to study the effects of ultraviolet radiation and NDP-MSH on the signalling proteins p38 and p53 *in vitro*. There has been increasing evidence for the involvement of these two proteins in pigmentary and ultraviolet radiation responses. p38 has been shown to be activated in response to α -melanocyte stimulating hormone in human melanocytes, and there was a synergistic interaction between ultraviolet radiation and melanocortin MC₁ receptor stimulation on p38 signalling (Newton et al., 2007). This may be important for the tanning response as ultraviolet radiation induced expression of proopiomelanocortin, tyrosinase and melanocortin MC₁ receptor in the melanocytes has been reported to be dependent on p38 mediated phosphorylation of the transcription factor USF-1 (Corre et al., 2004; Galibert et al., 2001). However, a recent report has also linked p38 with negative regulation of pigmentation via degradation of tyrosinase (Bellei et al., 2010). Ultraviolet radiation-induced DNA damage activates p53 in keratinocytes, which is able to induce the expression of the melanocortin precursor proopiomelanocortin (Cui et al., 2007). In the melanocytes, p53 is an early response gene induced by ultraviolet radiation, thought to be involved in DNA repair and cell

cycle arrest (Yang et al., 2006). p53 may also induce expression of the tyrosinase related genes (Nylander et al., 2000).

We wished to investigate the relative contributions of p38 and p53 signalling in response to ultraviolet radiation and NDP-MSH in co-cultures. All of our melanocyte strains have been genotyped for melanocortin MC₁ receptor so we can also look at the effect of melanocortin MC₁ receptor variant alleles. In these preliminary results, p38 was activated in melanocortin MC₁ receptor wild type co-cultures by ultraviolet radiation, with a synergistic effect of NDP-MSH (Fig. 4, lane 4). Melanocortin MC₁ receptor R151C co-cultures still activated p38 activation in response to ultraviolet radiation, however NDP-MSH no longer had a synergistic effect (Fig. 4, lane 8). In fact, the ultraviolet radiation p38 response seemed to be repressed in the presence of NDP-MSH in the R151C co-cultures. When compared with the results for monocultures (data not shown), it was concluded that the p53 activation was mainly due to the keratinocytes in the co-cultures. In contrast, the diminished responses of p38 in variant co-cultures might be attributed to the reduced function of the melanocortin MC₁ receptor variant receptor in the melanocytes.

These results confirm previous melanocyte monoculture studies in which phosphorylation of p38 was reduced in the red hair colour variants compared to the wild type melanocortin MC₁ receptor strains with NDP-MSH and ultraviolet irradiation (Newton et al., 2007). Using the co-culture model in combination with monoculture studies, we are able to delineate the contributions of the melanocortin MC₁ receptor signalling pathway to ultraviolet radiation and α -melanocyte stimulating hormone responses.

Dopachrome tautomerase is a member of the tyrosinase related protein family, and is also known as TYRP2. Dopachrome tautomerase has been recognized for its role in the formation of eumelanin by catalysing the conversion of dopachrome to 5,6-dihydroxyindole-2-carboxylic acid (DHICA) (Ito and Wakamatsu, 2008). Expression of dopachrome tautomerase is strongly induced by α -melanocyte stimulating hormone in melanocyte co-cultures (Roberts et al., 2008). The importance of dopachrome tautomerase beyond melanin synthesis has recently been realized with the finding that dopachrome tautomerase is able to promote cell survival and inhibit apoptosis in melanoma cells, possibly via inhibition of p53 (Sendoel et al., 2010). The biochemical mechanism of this inhibition is as yet unknown, but the authors speculated it might be related to the dopachrome tautomerase activity of this protein. Dopachrome tautomerase was also linked to DNA damage protection and reduced sensitivity to oxidative stress in amelanotic melanoma cells (Michard et al., 2008).

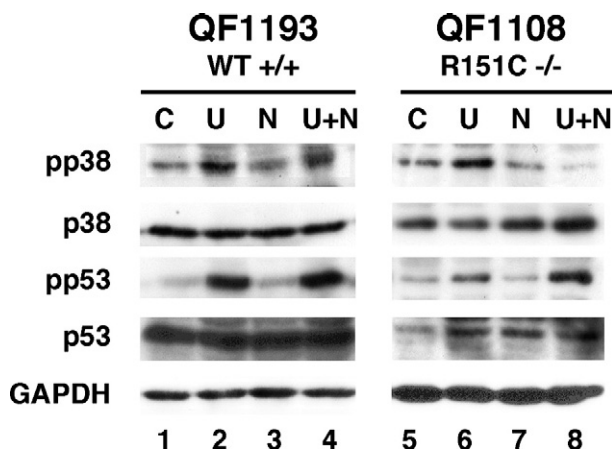


Fig. 4. Melanocyte co-culture p38 and p53 responses to NDP-MSH and ultraviolet radiation. Co-cultures of melanocortin MC₁ receptor wild type (WT – QF1193) or homozygous R151C melanocytes (QF1108) and keratinocytes were treated with 20 mJ/cm² ultraviolet radiation = U, 20 nM NDP-MSH = N or both = UN and incubated for 1 hr. Total cell extracts were used for western immuno-blotting. Activated forms of p38 and p53 were detected using antibodies specific for the Thr180/Tyr182 (pp38) and Ser15 (pp53) phosphorylated sites (Cell Signaling). These blots are representative of 2–3 independent experiments.

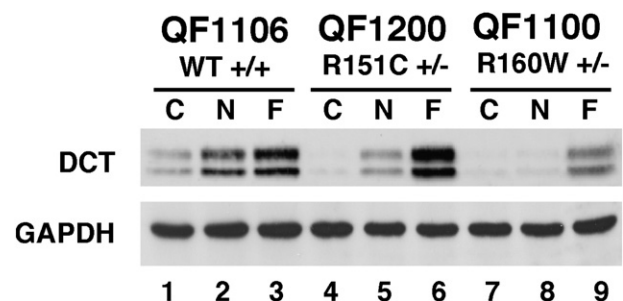


Fig. 5. Dopachrome tautomerase induction in melanocyte co-cultures. Melanocytes were either melanocortin MC₁ receptor wild type (WT – QF1106), heterozygous for R151C (QF1200) or heterozygous for R160W (QF1100). Co-cultures of melanocytes with keratinocytes were treated with the following compounds: C = Control, N = 20 nM NDP-MSH or F = 10 μM forskolin, for four days, then total cell extracts were used for western immuno-blotting with dopachrome tautomerase (Santa Cruz) and GAPDH antibodies. This blot is representative of responses seen in three different wild type and variant strains.

We have previously studied the induction of dopachrome tautomerase in melanocyte strains homozygous for melanocortin MC₁ receptor variants (Roberts et al., 2008). Here we have preliminary data on the induction of dopachrome tautomerase in heterozygote melanocortin MC₁ receptor strains in co-culture (Fig. 5). The wild type strain shows a good induction of dopachrome tautomerase by both NDP-MSH and forskolin (Fig. 5, lanes 2 and 3), whereas the variant strains, particularly the R160W heterozygote, show diminished responses to NDP-MSH (Fig. 5, lanes 5, 6, 8 and 9). These results are in line with our *in vitro* transfection studies in melanoma cells on the dominant negative effect of variant receptors on wild type melanocortin MC₁ receptor (Beaumont et al., 2007), and may also help explain the heterozygote effect on pigmentation phenotypes and skin cancer risk in genetic association studies.

5. Further models for studying melanocortin MC₁ receptor function

Although cell co-culture models allow interactions between melanocytes and keratinocytes, melanocytes integrated into the basal layer of a 3D reconstructed epidermis is closer to mimicking the *in vivo* situation in human skin, and has been used by a number of groups, particularly in the study of melanocyte responses to ultraviolet radiation (Duval et al., 2001). Immediate pigmentation can be seen in response to ultraviolet radiation, and it can be assumed that pigmentation could also be induced by melanocortin MC₁ receptor agonists, although the melanocortin MC₁ receptor has yet to be studied in the skin reconstruction model. One disadvantage of the 3D skin reconstruction is that cell and biochemical manipulations such as siRNA knockdown or overexpression studies are not easily performed. Imaging of cells also becomes more difficult with the move from a 2D to a 3D cell culture system.

Mouse models have been important for the identification of many different pigmentation genes via the study of coat colour mutants. Murine melanocortin MC₁ receptor maps to the mouse *extension* locus, which was known to influence pigmentation due to a number of naturally occurring dominant and recessive coat colour phenotypes. The *extension* locus alleles include wild type (E^+), recessive yellow (e), tobacco darkening (E^{tob}), sombre (E^{so}), and sombre-3J (E^{s-3J}) (Searle, 1968). The e allele encodes a non-functional mouse melanocortin MC₁ receptor, while the other dominant darkening alleles encode constitutively active or hyperactive mouse melanocortin MC₁ receptors (Robbins et al., 1993). Mutations in the melanocortin MC₁ receptor antagonist agouti signaling protein in mice also give rise to a number of coat colour phenotypes (Bultman et al., 1992). Temporal expression of agouti is responsible for the black/yellow banding pattern in wild type agouti mice, loss of agouti expression results in a black coat colour, whereas overexpression results in a yellow coat colour.

While we have discovered many important functions of the melanocortin system in mice, melanocortin MC₁ receptor regulated pigmentation in humans has some major differences. Agouti signaling protein does not produce the same obvious pheomelanin banding effect in human hair, indeed agouti signaling protein expression is yet to be detected in human skin, and the human melanocortin MC₁ receptor expressed in transgenic mice appeared resistant to the effects of agouti signaling protein (Healy et al., 2001). However, agouti signaling protein antagonism of the human melanocortin MC₁ receptor has been demonstrated *in vitro* (Suzuki et al., 1997). Moreover genome wide association studies have found associations between agouti signaling protein polymorphism and skin colour as well as skin cancer risk (Duffy et al., 2010), with the agouti signaling protein tagged rs4911442 risk allele showing epistatic effects roughly equivalent to a melanocortin MC₁ receptor r allele. Human melanocortin MC₁ receptor also has a higher sensitivity to ligands and a lower expression level relative to murine melanocortin MC₁ receptor. To address some of these issues, Jackson and co-workers developed mice

transgenic for human melanocortin MC₁ receptor, which was under control of the human melanocortin MC₁ receptor regulatory sequences (Jackson et al., 2007). In this mouse model, unlike the previous model where melanocortin MC₁ receptor was expressed under the control of the mouse regulatory sequences, agouti antagonism and a normal agouti banding pattern could be seen (Jackson et al., 2007).

Another major difference between mouse and human pigmentation is that mature mice do not have skin pigmentation, except for the ears and tail; this is due to a lack of melanocytes in the epidermis of the skin. D'Orazio and co-workers created a "humanized" mouse model, which mimics the human skin with the presence of melanocytes in the basal layer of the epidermis (D'Orazio et al., 2006). In this model, tanning in the trunk area of the transgenic melanocortin MC₁ receptor^{e/e} mouse could be achieved by topical application of the cAMP activating agent forskolin (D'Orazio et al., 2006).

Robinson and co-workers have created an albino and pigmented hairless melanocortin MC₁ receptor model in the mouse (Robinson et al., 2010). This allowed the authors to study the effects of melanocortin MC₁ receptor on not only pigmentation, but also non-pigmentary effects as well. The question of whether melanocortin MC₁ receptor really exerts an effect on skin cancer risk independent of pigmentation status seems to have been answered in this model. P53 positive keratinocyte clones were used as a readout for inadequate repair of DNA damage and clonal proliferation. Significantly higher numbers of p53-positive clones were found in albino mice null for melanocortin MC₁ receptor, compared to albino mice expressing wild type melanocortin MC₁ receptor. This indicates that melanocortin MC₁ receptor, at least in this model, does have non-pigmentary protective effects on skin cancer development.

Another important tool used in the study of melanocortin MC₁ receptor function is microarray analysis. It is becoming increasingly clear the melanocortin MC₁ receptor signalling activates a large number of target genes involved in many processes such as melanin synthesis, melanosome granule maturation, melanosome trafficking and transfer, cell morphology, DNA repair, cell survival and others. A recent study identified 255 genes that were altered by α -melanocyte stimulating hormone in murine melanocytes (Le Pape et al., 2009). Unsurprisingly, α -melanocyte stimulating hormone treatment upregulated known genes important for pigmentation, while murine agouti signaling protein, as an inverse agonist (Siegrist et al., 1997), downregulated the same genes. Surprisingly, murine agouti signaling protein also upregulated a number of additional genes involved in differentiation and developmental processes, as well as cell adhesion, motility and extra cellular matrix–receptor interactions (Le Pape et al., 2009). The authors suggested that agouti signaling protein activates expression of genes that are typically expressed during morphogenesis, and are reactivated in cancer cells to promote migration and invasion. Supporting this, melan-a melanocytes and B16 melanoma cells treated with murine agouti signaling protein showed increased migration.

6. Conclusions

The use of naturally occurring mouse coat colour mutants, human genetic association studies, *in vitro* studies using transfected cells or genetically defined melanocyte monocultures and co-cultures have all contributed to our knowledge of melanocortin MC₁ receptor function in human pigmentation and skin cancer risk. It is becoming increasingly apparent that the function of melanocortin MC₁ receptor is much more complex than just regulating the switch from the red/yellow pheomelanin to the brown/black eumelanin that was originally described. Future melanocortin MC₁ receptor pharmacogenetic studies may utilise new transgenic mouse models, new

information about novel melanocortin MC₁ receptor response genes obtained in microarray studies, or more physiologically relevant models of the human skin such as *in vitro* 3D reconstructions.

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